

Naturally Occurring Rhamnopyranose Esters as Anticancer Agents: Molecular Docking and ADMET Study

Md. Inshaful Islam^{*,a}, Sulaiman Bin Sultan^a, Takbir Hossain^a, Md. Mohin Hasnain^a, Md. Badrul Islam^b and Abul K.M.S. Azad^b

^a Department of Chemistry, Faculty of Science, University of Chittagong, Chittagong, 4331, Bangladesh

^b Department of Chemistry, Chattogram Govt. College, Chittagong, 4203, Bangladesh

Abstract

After heart disease, cancer continues to be the second most prevalent cause of death in the USA. Several chemotherapeutic treatments (drugs) are available for cancer that use powerful chemicals to kill the body's rapidly proliferating cells. However, recent research disclosed that many clinically viable anticancer drugs have been developed with the help of chemicals originating from plants. A number of phytochemicals isolated from plants possess rhamnopyranoses and some of them are acyl rhamnopyranoses. Encouragingly, such compounds were reported for their cell proliferation and migration inhibition activities against invasive human triple-negative breast cancer cells. In this study, four naturally occurring rhamnopyranose esters were checked against three cancer-related proteins (PDB IDs: 3TJM, 4OAR, and 5FGK) via molecular docking. Rhamnose compounds 3-6 showed better binding energy compared to the related standard drugs in use in the hospitals. Compound 6 was found highly potential against all the proteins (-8.5 to -11.3 kcal/mol). ADMET studies have also been discussed in this respect. This study indicated that natural rhamnopyranose esters could be used to stop the spreading of cancer cells like other reported sugar fatty acid esters (SFAEs).

Keywords: Anticancer drugs, Cancer, Molecular docking, Natural products, Phytochemicals, Sugar esters, Sugar fatty acid esters (SFAEs).

1. Introduction

Cancer is a condition marked by abnormal cell growth. It is regarded as one of the world's deadliest diseases. Men are more likely to develop prostate, lung, stomach, colorectal, and liver cancers than women, who are more likely to develop colorectal, breast, lung, thyroid, and cervical cancers [1]. In fact, according to the WHO, cancer is a broad category of disease that can begin in virtually every organ or tissue of the body when aberrant cells grow out of control [1]. By altering or eliminating significant risk factors and putting into practice currently recommended evidence-based preventative methods, between 30% and 50% of cancer-related fatalities could be avoided. However, once a person is affected by uncontrolled cell growth medication/therapeutic treatment is essential. Standard chemotherapy does not always stop cell motility; instead, it generally causes programmed cell death in cancer cells that divide quickly through mechanisms such as suppression of cell division

* Corresponding author. Tel.: +880 1643 947406
E-mail address: mdinshafulislam@gmail.com

Manuscript History:

Received 08 October, 2022, Revised 18 October, 2022, Accepted 18 October, 2022, Published 31 October, 2022
Copyright © 2022 UNIMAS Publisher. This is an open access article under the CC BY-NC-SA 4.0 license.
<https://doi.org/10.33736/jaspe.5065.2022>

and interruption of deoxyribonucleic acid/ribonucleic acid (DNA/RNA) synthesis [2]. Therefore, searching for novel therapeutic agents is essential.

There is a growing tendency to use natural compounds as drugs and some of them play an important role in the discovery of anticancer antioxidants [3,4]. For instance, medications used in clinical settings such as taxanes, doxorubicin, epothilones, vincristine, and camptothecin are either derived from natural products or were discovered from parent natural product compounds [5,6]. Carbohydrates are abundant in phytochemicals and sugar esters (SEs) are also present in them. Sugar fatty acid esters (SFAEs) are biodegradable, non-ionic surfactants with amphiphilic properties of interest to prepare many biologically active and household products [7]. They have broad uses in the food, pharmaceutical, detergent, agricultural, fine chemical, and personal care sectors because of their effective stabilizing and conditioning qualities [8-13]. SFAEs can be created through the esterification of non-polar fatty acids with sugar/sugar alcohols [14-20]. Their properties vary with the degree of substitution (DS) and hence, they have attracted a lot of scientific attention. Moreover, having numerous stereocenters and functionality, carbohydrates provide synthetic chemists a convenient way to obtain chiral intermediates at low cost for application in the synthesis of natural products [21,22].

Among the monosaccharides, L-rhamnopyranose is a widely distributed natural sugar that is present in a wide variety of natural carbohydrates [23,24]. For instance, the actinomycete *Norcardia brasiliensis* IFM0406 broth included brasilicardin A (**1**, Figure 1), which has two rhamnopyranosyl moieties. The immunosuppressive properties are brought on by the addition of 3-hydroxybenzoate esters in the 3- and 4-positions of its rhamnose unit(s) [25]. Against glutamate-induced neurotoxicity, 2-O-acetyl-3,4-di-O-(*E*)-p-methoxycinnamoyl-L-rhamnopyranoside from *S. buergeriana* showed significant protective benefits [26]. The enzyme 3-hydroxysteroiddehydrogenase, which is a useful target in the search for antiphlogistic and antitumor drugs, has been reported to be inhibited by a number of rhamnopyranoside derivatives that have been isolated and discovered from several species of *Streptomyces* [27,28]. In our Organic laboratory, several rhamnose esters (**2a-e**) were synthesized and found to show antimicrobial potentiality [29-33].

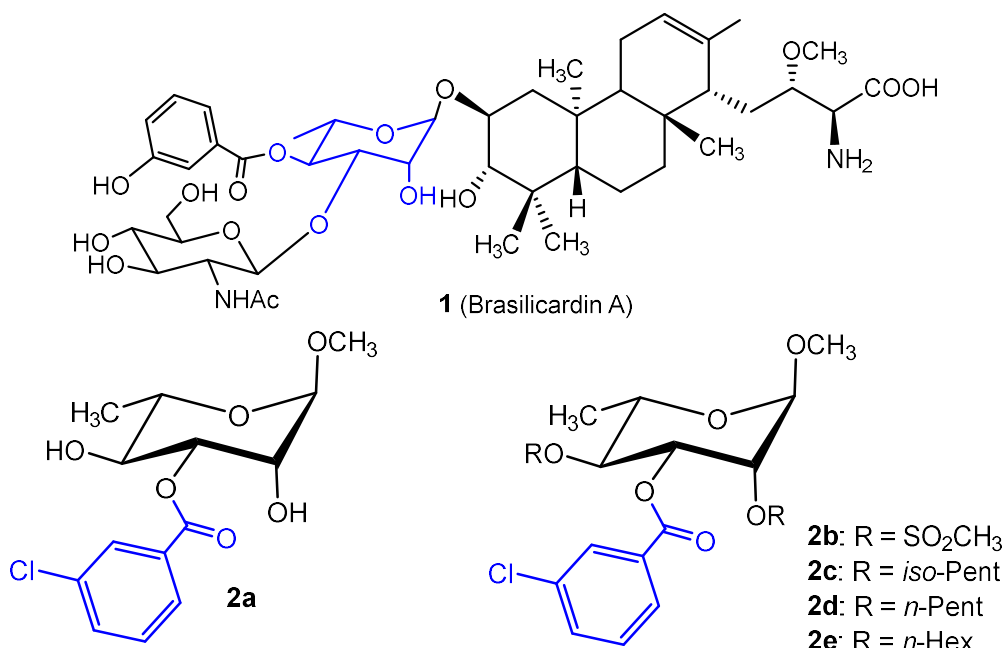


Figure 1. Natural and synthetic rhamnose compounds of biological interest.

There is a long tradition of using medicinal herbs to treat various disorders, including cancer. Several clinically viable anticancer drugs have been developed with the help of chemicals originating

from plants. In this context, some compounds (quercetin, quercitrin, and salanin, etc.) from the leaves of *Azadirachta indica* were performed docking studies with yeast Cdk protein and showed good interaction with the cdc28 protein [34]. Recently (2022), ethyl acetate and chloroform extracts of *Ifloga spicata* were found highly active against HepG-2 cell line (MTT assay, IC₅₀ values 5.54 and 6.52 µg/mL, respectively) [35]. According to Snoch et al. [36], altering the hydrophobic chains of sugar fatty acid esters (SFAEs) can change how hazardous they are to human skin melanoma and prostate cancer cell lines. The synthesized SFAEs showed potential metastatic properties with IC₅₀ values 63.3–1737.6 µM.

As a continuation of our research work on bioactive molecules [37-41], we were interested to check their anticancer potentiality. Thus, four natural rhamnopyranoside esters are considered for their anticancer properties *via* molecular docking and ADMET (absorption, distribution, metabolism, excretion, and toxicity) studies.

2. Materials and methods

2.1. Natural rhamnose-based SFAEs

As mentioned earlier, rhamnose-based esters are available in natural products [42,43]. Compound **3** was isolated and identified from the *Streptomyces* sp. strain TN58 found in Tunisian soil [44]. In 2020, Elmaidomy and co-workers [45] successfully isolated several phytochemicals from the stems of *Premna odorata* B. Among them, three rhamnopyranose-based esters **4-6** were characterized by spectral methods. Tests for antioxidant and cell proliferation inhibition activities (using human triple-negative breast cancer cells) were conducted. The results were unambiguously positive and showed enhanced biological potentiality of the separated compounds [45]. These compounds **3-6** are used in the present study.

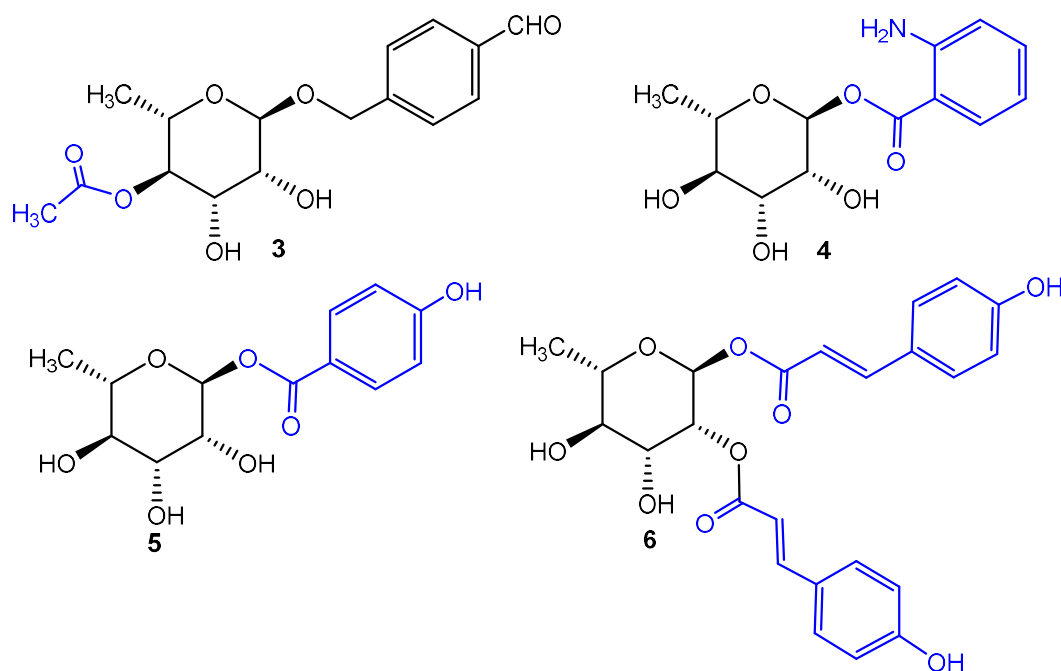


Figure 2. Structures of natural rhamnopyranoside-based SEs 3-6 for anticancer study.

2.2. Methods used for molecular docking

Computer-aided drug design typically makes use of molecular docking (CADD). It can be used

at many stages of the drug design process to: (i) anticipate the binding mode of already known ligands; (ii) find novel and potent ligands; and (iii) as a tool to forecast binding affinity [46,47]. In our study, we used molecular docking of compounds **3-6** employing freely available PyRx (AutoDock) software [48,49].

Ligand preparation: Firstly correct structures of all the natural rhamnoses **3-6** are drawn in ChemDraw 18.0 and saved as MOL file. These are opened in Chem3D and minimized with MM-2 followed by saving as SDF formats and used for docking in the next step.

Protein preparation: Three cancer-related proteins such as (i) human fatty acid synthase thioesterase (PDB: 3TJM), (ii) progesterone receptor peptide (PDB: 4OAR), and (iii) cyclin-dependent kinase (CDK8) (PDB: 5FGK) were retrieved from the popular RCSB Protein Data Bank [50,51]. After opening these in Discovery Studio, these are dried and unnecessary atoms are removed. These are further opened in Swisspdb software, conducted energy minimization, and saved as pdbqt format.

Docking: In the PyRx software (AutoDock), proteins are loaded followed by the import of ligands. The ligands **3-6** are further subjected for energy minimization with PyRx-associated software. At this stage, ligands are selected and proceed to the next step where maximum box sizes were used. Docked complexes were duly saved, and opened in the Discovery Studio and necessary data has been collected [52,53].

2.3. ADMET calculation

Chemical absorption, distribution, metabolism, excretion, and toxicity (ADMET) profiling aids in reducing potential dangers throughout clinical development [54]. Scientists are aware that ADMET profiling is required to identify whether or not a molecule is suitable to advance to the clinical stage because drug development failures might be caused by efficacy or safety difficulties [55,56]. Utilizing the pkCSM tool (<http://biosig.unimelb.edu.au>), *in silico* ADMET was calculated for the current experiment [57]. For the calculation of pkCSM-pharmacokinetics, the appropriate stereochemical structures of the SEs **3-6** were translated to the respective SMILES (simplified molecular-input line-entry system) and SD file formats. Using SwissADME's free web tools (<http://www.swissadme.ch>), the comparable SMILES and SD file formats were also employed to predict drug-likeness features [58].

3. Results and discussion

3.1. Molecular docking: Binding energy with cancer causing protein 3TJM

Mixed, targeted, and customized therapy is being employed to treat cancer increasingly frequently these days. The most promising treatments target tumor cells in a host body using inspiration from nature. Synthetic and highly effective therapies sometimes look insufficient as well as costly. Attention was made to get natural therapeutic agents like sugar fatty acids esters (SFAEs) and related research started in 1970 [59]. Experimental results explicit SFAEs' ability to inhibit excessive proliferation of cells in the bone marrow. Thus, in the present study, four natural SFAEs **3-6** are considered and checked for their ability to anti-proliferation of cells against three cancer-related proteins.

Firstly, we considered human fatty acid synthase (hFAS) (PDB: 3TJM) as it catalyzes a variety of fatty acid synthesis in the body and causes several diseases including cancers. Hence, hFAS is a target for anti-cancer drug development [60]. Molecular docking binding affinities of **3-6** with 3TJM are shown in Table 1. Binding affinities of the natural SFAEs (-6.3 to -11.3 kcal/mol) are comparable to the standard anticancer drug topotecan (-7.4 kcal/mol). Unsaturated acyl rhamnopyranose ester **6** showed the highest binding affinity (-11.3 kcal/mol) followed by 3-*O*-acetyl-rhamnose **3** (-7.7

kcal/mol). Rhamnopyranose-based SFAEs **3-6** showed several interactions with amino acid (AA) residues of the enzyme 3TJM like standard topotecan (Table 2). For better visualization, interactions between SFAEs and enzymes are also presented in Figure 3.

Table 1. Molecular docking score (binding affinity) with 3TJM.

Ligand/ Drug	Binding affinity (kcal/mol)	No. of H bond	No. of Hydrophobic bond	No. of van der Waal/ES bond ^a	Total bonds
3	-7.7	2	5	0	7
4	-6.3	0	6	0	6
5	-6.5	0	6	0	6
6	-11.3	0	8	0	8
Topotecan (TPT)	-7.4	3	5	2	10

^aES = electrostatic; so far -6.0 kcal/mol is standard docking score

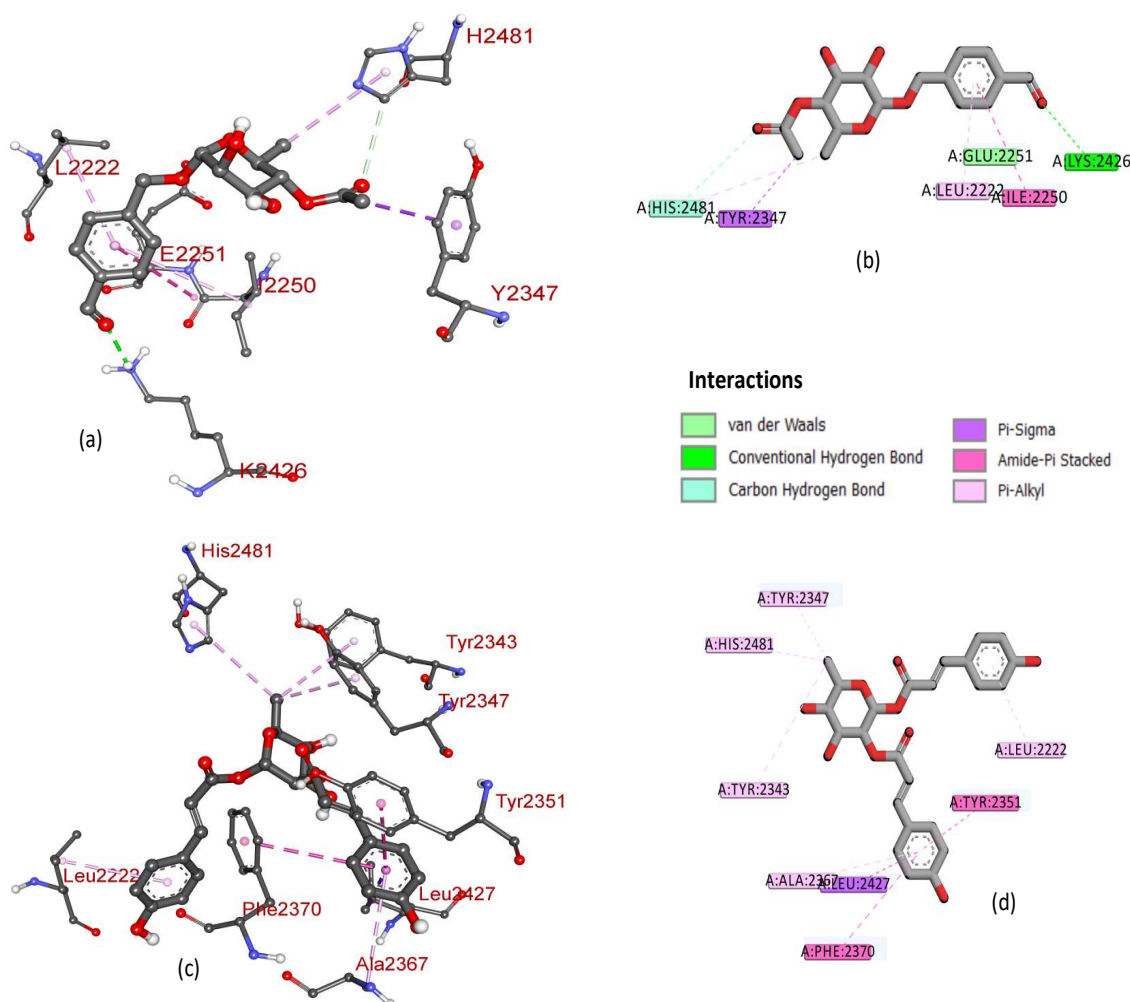


Figure 3. 3D and 2D non-bond interactions of 3TJM with ligands - (a), (b) **3**; and (c), (d) **6**.

Table 2. Interaction of compounds with amino acid (AA) residues in compounds-3TJM complex.

Drug	Hydrogen bond		Hydrophobic bond		Van der Waals bond	
	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)
3	LYS2426 HIS2481	2.30659 3.60039	TYR2347	3.77739	Absent	
			ILE2250	4.81434		
			HIS2481	4.87778		
			LEU2222	4.68179		
			ILE2250	5.37677		
4			TYR2347	4.44896	Absent	
			ILE2250	4.46877		
			PHE2370	5.47883		
			PHE2423	4.42897		
			ILE2250	4.70364		
5			TYR2347	4.44896	Absent	
			ILE2250	4.46877		
			PHE2370	5.47883		
			PHE2423	4.42897		
			ILE2250	4.70364		
6			LEU2427	3.89325	Absent	
			TYR2351	4.10401		
			PHE2370	5.72170		
			TYR2343	5.31378		
			TYR2347	4.18470		
Topotecan (TPT)	SER2221 GLU2251 TYR2307	2.59638 2.91400 3.14200	ALA2482	4.63781	GLU2251 GLU2251	3.51756 4.07741
			HIS2481	5.17995		
			VAL2256	5.37410		
			ALA2482	4.67044		
			ILE2250	5.18481		

Note: TRP = Tryptophan, ASP = Aspartic acid, GLU = Glutamic acid, LEU = Leucine, THR = Threonine, ASN = Asparagine, GLN = Glutamine, PHE = Phenylalanine, ILE = Isoleucine, ARG = Arginine, VAL = Valine, SER = Serine, PRO = Proline, GLY = Glycine, HIS = Histidine, LYS = Lysine, TRP = TRPosine, CYS = Cysteine, MET = Methionine.

3.2. Molecular docking: Binding energy with cancer causing protein 4OAR

Our next target enzyme was the human progesterone receptor (PR) related protein (PDB: 4OAR). Most breast cancer drugs are targeting human PR [61]. For example, a highly selective PR antagonist is ulipristal acetate (UPA) is available in the market. Thus, 4OAR was adopted for molecular docking with rhamnose-based SEs **3-6**. Docking results are presented in Table 3.

It is clear from Table 3 that SEs **3-6** have a higher binding affinity (BA) (-7.7 to 8.9 kcal/mol) compared to the standard ulipristal acetate (-6.9 kcal/mol). Rhamnose ester **6** showed the maximum BA (-8.9 kcal/mol) followed by acetate **3** (-8.5 kcal/mol). All the compounds have a variety of bonding and non-bonding interactions compared to the ulipristal acetate (Table 4 and Figure 4). Both ulipristal acetate and compound **6** interact with CYS891 in chain A. Additionally, rhamnose esters several hydrogen bonds with the amino acid residues of chain A (Table 4). These H-bonds are

probably responsible for their higher binding affinities.

Table 3. Molecular docking score (binding energy) with 4OAR.

Ligand/ Drug	Binding energy (kcal/mol)	No. of H bond	No. of Hydrophobic bond	No. of van der Waal/ES ^a bond	Total bonds
3	-8.5	3	4	1	8
4	-7.8	6	6	1	13
5	-7.7	4	6	1	11
6	-8.9	3	7	0	10
Ulipristal acetate	-6.9	0	2	0	2

^aES = electrostatic; so far -6.0 kcal/mol is standard docking score

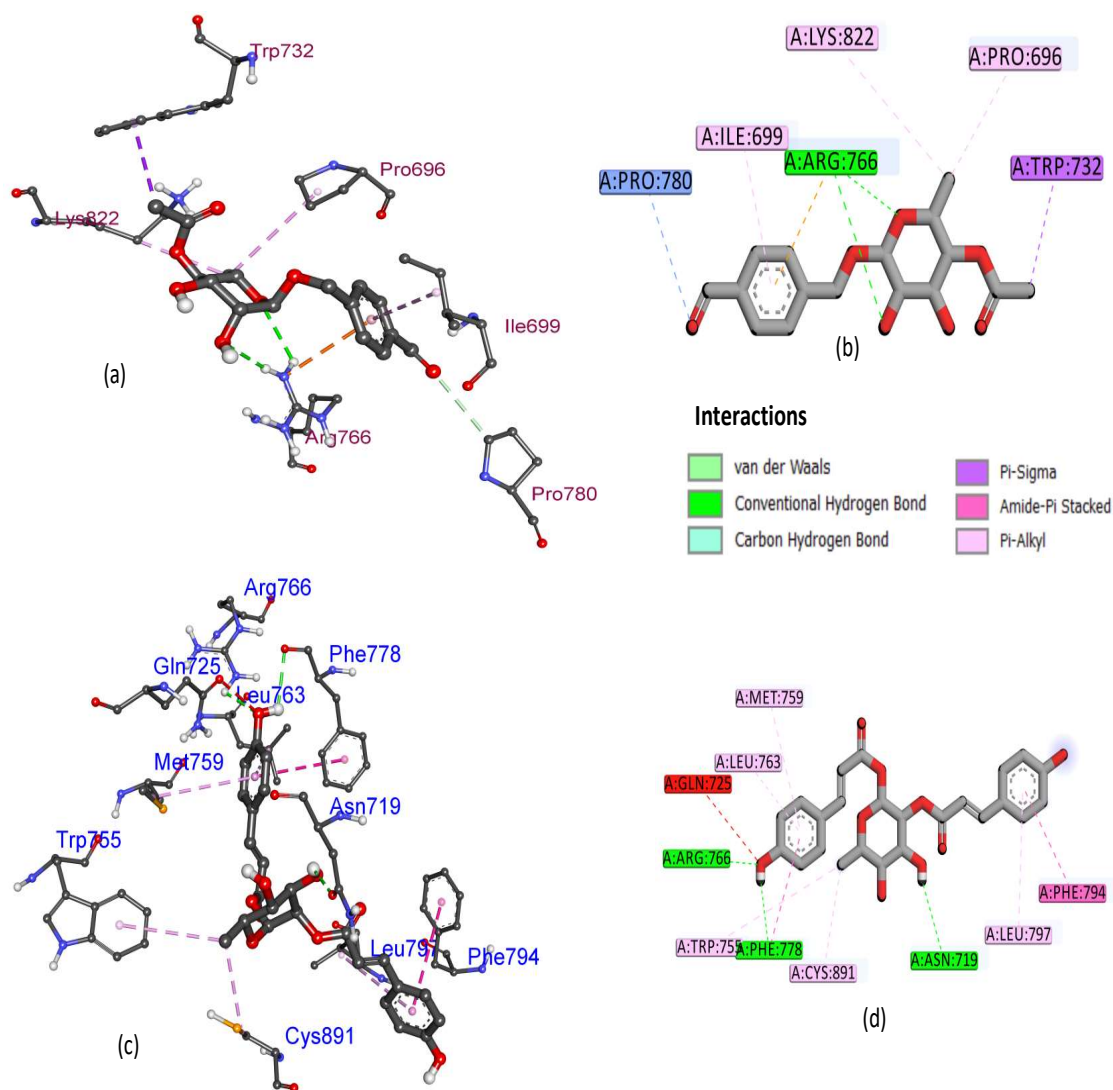


Figure 4. 3D and 2D non-bond interactions of 4OAR with ligands - (a), (b) 3; and (c), (d) 6.

Table 4. Interaction of compounds with amino acid (AA) residues in compounds-4OAR complex.

Drug	Hydrogen bond		Hydrophobic bond		Van der Waals /ES bond	
	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)
3	ARG766	2.71903	TRP732	3.98838	ARG766	4.02901
	ARG766	2.94511	PRO696	4.95069		
	PRO780	3.50793	LYS822	5.23245		
			ILE699	4.58957		
4	ARG766	2.20714	VAL698	3.87101	ARG766	4.21398
	ILE699	2.88715	ASP697;	4.64534		
	PHE778	2.57122	VAL698	4.97147		
	GLN725	2.19084	PRO696	5.17104		
	UNK1	2.34771	LYS822	5.24640		
	ILE699	2.63355	PHE818	5.27371		
5	SER728	2.83544	VAL729	3.60331	LYS822	3.24195
	ARG766	2.04286	TRP732	5.57146		
	PRO696	2.41457	TRP732	5.52667		
	LYS822	3.24195	ILE699	4.24415		
			PRO696	4.83511		
			LEU758	5.30345		
6	ARG766	2.49600	PHE794	5.12810	Absent	
	PHE778	2.64071	PHE778	4.99925		
	ASN719	2.36963	CYS891	4.10214		
			TRP755	5.01043		
			MET759	5.15711		
			LEU763	5.10761		
			LEU797	5.41374		
Ulipristal acetate			MET759	4.60149	Absent	
			CYS891	4.87029		

Note: TRP = Tryptophan, ASP = Aspartic acid, GLU = Glutamic acid, LEU = Leucine, THR = Threonine, ASN = Asparagine, GLN = Glutamine, PHE = Phenylalanine, ILE = Isoleucine, ARG = Arginine, VAL = Valine, SER = Serine, PRO = Proline, GLY = Glycine, HIS = Histidine, LYS = Lysine, TRP = TRPosine, CYS = Cysteine, MET = Methionine.

3.3. Molecular docking: Binding energy with cancer causing protein 5FGK

With good molecular BAs of rhamnopyranose SEs **3-6** in the previous sections, we employed another cancer-related enzyme cyclin-dependent kinase (CDK8; PDB: 5FGK). In colon and stomach malignancies, the expression of the CDK8 gene is correlated with the activation of -catenin, a crucial transcriptional regulator of canonical WNT signaling [62]. Molecular docking BAs of **3-6** with 5FGK are presented in Table 5. In this case also, compound **3** and **6** showed excellent binding affinities. For validation, docking scores are compared with standard drug named ribociclib. Ribociclib (KISQALI[®]) is a pill that is referred to as a targeted treatment because it interferes with or blocks chemicals that are involved in the proliferation of cancer cells. It is not chemotherapy. With respect to ribociclib (-8.9 kcal/mol) rhamnose esters showed slightly lower binding affinities (-8.5 to -7.5 kcal/mol). For better understanding, related interactions are presented in Figure 5 and Table 6 including standard drug.

Table 5. Molecular docking score (binding energy) with 5FGK.

Ligand/ Drug	Binding energy (kcal/mol)	No. of H bond	No. of Hydrophobic bond	No. of van der Waal/ES ^a bond	Total bonds
3	-7.7	3	5	0	8
4	-7.6	5	5	0	10
5	-7.5	4	4	0	8
6	-8.5	4	8	0	12
Ribociclib	-8.9	2	5	0	7

^aES = electrostatic; so far -6.0 kcal/mol is standard docking score

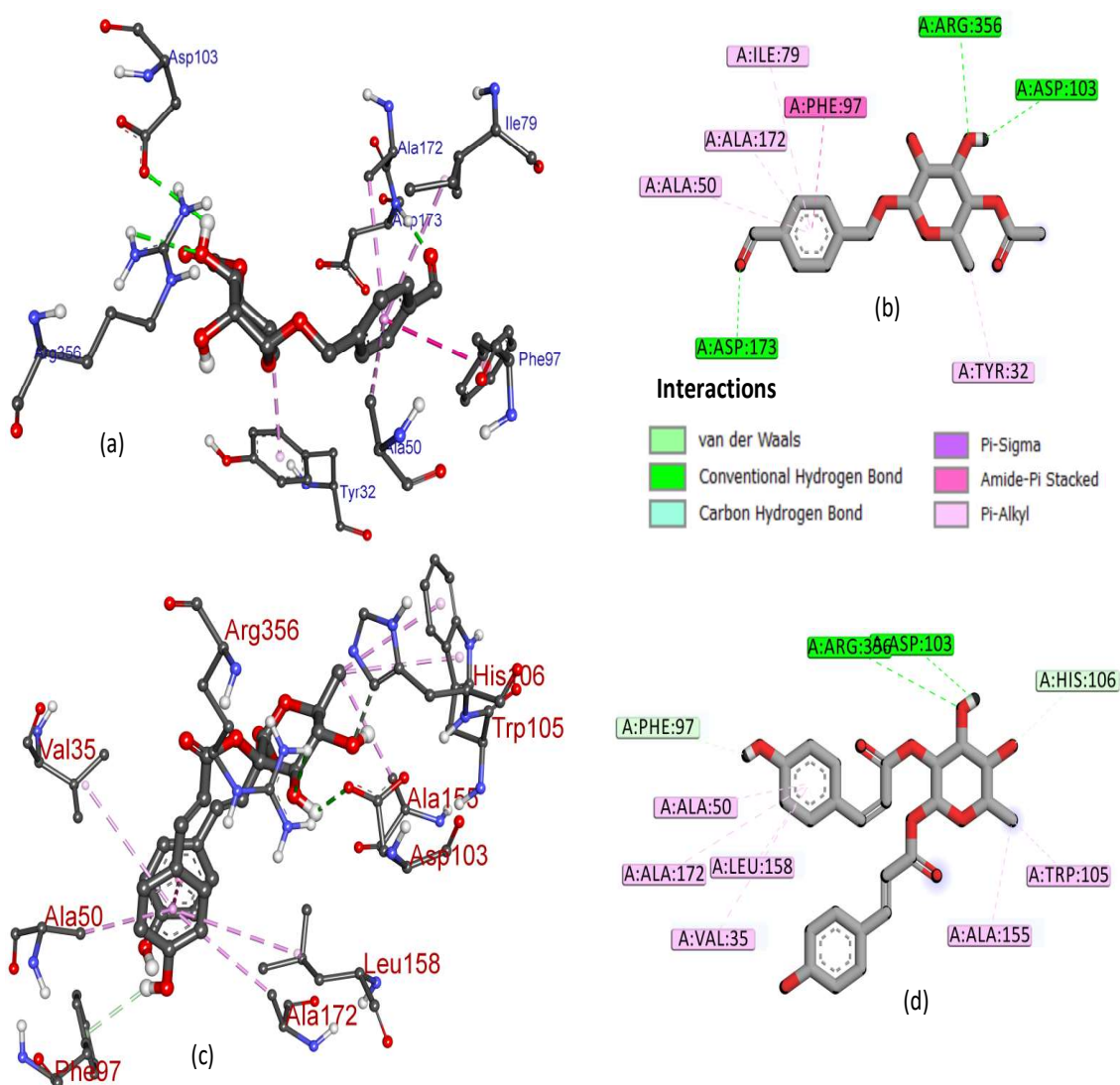


Figure 5. 3D and 2D non-bond interactions of 5FGK with ligands - (a), (b) 3; and (c), (d) 6.

Table 6. Interaction of compounds with amino acid (AA) residues in compounds-5FGK complex.

Drug	Hydrogen bond		Hydrophobic bond		Van der Waals /ES bond	
	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)	Interacting residue of amino acid	Distance (Å)
3	ASP173	1.95773	PHE97	4.11637	Absent	
	ARG356	2.96779	TYR32	4.01255		
	ASP103	2.87197	ALA50	5.30722		
			ILE79	5.07992		
4	ALA100	2.79575	ALA172	4.47971	Absent	
	ARG356	3.01603	ILE79	3.96125		
	UNK1	2.78748	LEU158	5.18217		
	ASP98	2.52265	PHE97	4.56786		
	TYR99	3.30698				
5	ARG356	2.96723	TYR32	3.69290	Absent	
	ARG356	2.33521	ALA172	3.72080		
	ALA100	2.35948	LEU158	4.69089		
	ASP98	2.84600	VAL35	4.94948		
6	ARG356	2.85220	UNK1	3.68384	Absent	
	ASP103	2.39185	ALA155	4.39449		
	HIS106	3.74545	TRP105	4.81519		
	PHE97	2.91772	TRP105	4.72126		
			VAL35	5.22754		
			ALA50	4.67866		
			LEU158	5.40142		
		ALA172	5.25045			
Ribociclib	LYS52	2.27737	TYR32	3.90735	Absent	
	GLU357	2.59651	VAL35	3.89390		
			ALA50	5.29917		
			LYS52	4.69478		
			TYR32	5.27449		

Note: TRP = Tryptophan, ASP = Aspartic acid, GLU = Glutamic acid, LEU = Leucine, THR = Threonine, ASN = Asparagine, GLN = Glutamine, PHE = Phenylalanine, ILE = Isoleucine, ARG = Arginine, VAL = Valine, SER = Serine, PRO = Proline, GLY = Glycine, HIS = Histidine, LYS = Lysine, TRP = TRPosine, CYS = Cysteine, MET = Methionine.

3.4. ADMET results of rhamnopyranose esters 3-6

Having good molecular binding affinities rhamnopyranose SEs **3-6** are placed for ADMET calculations. For validity, three standard anticancer drugs are also used for ADMET prediction. As shown in Table 7, human intestinal absorption (HIA) and p-glycoprotein inhibition (P-gpi) of the SEs are comparable to the standard anticancer drugs (topotecan, ribociclib). All the compounds can pass through the blood-brain barrier (BBB) and central nervous system. However, they are non-substrate of CYP3A4 (except **3**). Their excretion in terms of total renal clearance is comparable to the standard drugs (Table 7). Encouragingly, these compounds **3-6** are non-inhibitor of hERG and hence these SEs are safe. These ADME data are in accord with previously reported articles [63.64].

Table 7. ADMET properties of rhamnose compounds 3-6 with standard drugs.

Drug	Absorption			Distribution		Metabolism	Excretion	Toxicity	
	C2P	HIA (%)	P-gpI	BBB	CNS			CYP3A4 substrate	Total clearance (mL/min/kg)
				(permeability)					
3	1.323	95.751	Yes	-1.234	-3.237	Yes	0.698	No	2.585
4	0.210	56.289	No	-0.807	-3.632	No	0.221	No	1.616
5	0.255	53.035	No	-0.972	-3.179	No	0.629	No	1.960
6	-0.243	58.757	Yes	-1.343	-3.392	No	0.219	No	3.036
Topotecan	0.926	76.772	No	-1.005	-3.823	Yes	1.144	No	3.061
Ulipristal acetate	0.706	98.895	Yes	-0.038	-1.684	Yes	0.249	No	2.706
Ribociclib	1.323	95.751	Yes	-1.234	-3.237	Yes	0.698	No	2.585

4. Conclusion

Natural phytochemicals serve as vital resources for developing new medications and are also used in the treatment of cancer. Here, 4 natural rhamnopyranose esters **3-6** are successfully docked with cancer-causing three enzymes, namely human fatty acid synthase (3TJM), human progesterone receptor (4OAR), and cyclin-dependent kinase (5FGK). The results are found to be encouraging and are in conformity with some *in vitro* results. Especially, 2-*O*-(*p*-hydroxycinnamoyl)-rhamnose ester **6** and 4-*O*-ehanoylrhamnose ester **3** bearded excellent binding affinities with all three types of cancer target proteins with a good number of hydrogen bonds. These observations will hopefully pave a way for the establishment of anticancer agents from the biodegradable rhamnopyranose scaffold.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

References

- [1] WHO Cancer, (2022). Available online: <https://www.who.int/health-topics/cancer> (accessed on 2 October, 2022).
- [2] Giovannucci, E., Harlan, D.M., Archer, M.C., Bergenstal, R.M., Gapstur, S.M., Habel, L.A., Pollak, M., Regensteiner, J.G., & Yee, D. (2010). Diabetes and cancer: a consensus report. *Ca-Cancer J. Clin.*, 60, 207–221. <https://doi.org/10.3322/caac.20078>.
- [3] Leporini, M., Catinella, G., Bruno, M., Falco, T., TundisR., & Loizzo, M.R. (2018). Investigating the antiproliferative and antioxidant properties of *Pancreaticum maritimum* L. (Amaryllidaceae) stems, flowers, bulbs, and fruits extracts. *J. Evidence-Based Complementary Altern. Med.*, 2018, 1–8. <https://doi.org/10.1155/2018/9301247>
- [4] Sharma, N., Sharma, A., Bhatia, G., Landi, M., Brestic, M., Singh, B., Singh, J., Kaur, S., & Bhardwaj, R. (2019). Isolation of phytochemicals from *Bauhinia variegata* l. Bark and their *in vitro* antioxidant and cytotoxic potential. *Antioxidants*, 2019(8), 492. <https://doi.org/10.3390/antiox8100492>
- [5] Khan, T., & Gurav, P. (2018). Phyto nanotechnology: Enhancing delivery of plant based anticancer drugs. *Frontiers in Pharmacology*, 8, 1002. <https://doi.org/10.3389/fphar.2017.01002>
- [6] Cragg, G.M., & Pezzuto, J.M., (2016). Natural products as a vital source for the discovery of cancer chemotherapeutic and chemopreventive agents. *Medical Principles and Practice*, 25, 41-59.

- <https://doi.org/10.1159/000443404>
- [7] Koumba Ibinga, S.K., Fabre, J.-F., Bikanga, R., & Mouloungui, Z. (2019). Atypical reaction media and organized systems for the synthesis of low-substitution sugar esters. *Front. Chem.*, 7, 587. <https://doi.org/10.3389/fchem.2019.00587>
- [8] Rahman, M.A., Chakma, U, Kumer, A., Rahman, M.R., & Matin, M.M. (2023). Uridine-derived 4-aminophenyl 1-thioglucosides: DFT optimized FMO, ADME, and antiviral activities study. *Biointerface Research in Applied Chemistry*, 13(1), 52. <https://doi.org/10.33263/BRIAC131.052>
- [9] Matin, M.M., Bhuiyan, M.M.H., Azad, A.K.M.S., & Rashid, M.H.O. (2015). Synthesis of 6-O-stearoyl-1,2-O-isopropylidene- α -D-glucopyranose derivatives for antimicrobial evaluation. *Journal of Physical Science*, 26(1), 1-12. ISSN: 2180-4230
- [10] El-Baz, H.A., Elazzazy, A.M., Saleh, T.S., Dourou, M., Mahyoub, J.A., Baeshen, M.N., Madian, H.R., & Aggelis, G. (2021). Enzymatic synthesis of glucose fatty acid esters using SCOs as acyl group-donors and their biological activities. *Appl. Sci.*, 11, 2700. <https://doi.org/10.3390/app11062700>
- [11] Kabir, A.K.M.S., Matin, M.M., Hossain, A., & Sattar, M.A. (2003). Synthesis and antimicrobial activities of some rhamno-pyranoside derivatives. *Journal of the Bangladesh Chemical Society*, 16(2), 85–93. ISSN: 1022-016X
- [12] Matin, M.M., & Chakraborty, P. (2020). Synthesis, spectral and DFT characterization, PASS prediction, antimicrobial, and ADMET studies of some novel mannopyranoside esters. *Journal of Applied Science & Process Engineering*, 7(2), 572–586. <https://doi.org/10.33736/jaspe.2603.2020>
- [13] Kabir, A.K.M.S., Matin, M.M., Mridha, M.A.U., & Shahed, S.M. (1998). Antifungal activities of some methyl 6-O-trityl- α -D-mannopyranosides. *The Chittagong University Journal of Science*, 22(1), 41–46. ISSN: 1561-1167
- [14] Kabir, A.K.M.S., Matin, M.M., & Uddin, M.R. (1998). Comparative studies on selective acylation of uridine using the dibutyltin oxide and direct methods. *The Chittagong University Journal of Science*, 22(1), 97-103. ISSN: 1561-1167
- [15] Yang, Z., & Huang, Z.-L. (2012). Enzymatic synthesis of sugar fatty acid esters in ionic liquids. *Catalysis Science & Technology*, 2, 1767-1775. <http://dx.doi.org/10.1039/C2CY20109G>
- [16] Islam, N, Islam, M.D., Rahman, M.R., & Matin, M.M. (2021). Octyl 6-O-hexanoyl- β -D-glucopyranosides: Synthesis, PASS, antibacterial, in silico ADMET, and DFT studies. *Current Chemistry Letters*, 10(4), 413–426. <http://dx.doi.org/10.5267/j.ccl.2021.5.003>
- [17] Matin, M.M. (2008). Synthesis of D-glucose derived oxetane: 1,2-O-isopropylidene-4-(S)-3-O,4-C-methylene-5-O-methanesulfonyl- β -L-threo-pento-1,4-furanose. *Journal of Applied Sciences Research*, 4(11), 1478–1482. ISSN: 1816-157X
- [18] Matin, M.M., Bhuiyan, M.M.H., Hossain, M.M., & Roshid M.H.O. (2015). Synthesis and comparative antibacterial studies of some benzylidene monosaccharide benzoates. *Journal of the Turkish Chemical Society Section A: Chemistry*, 2(4), 12–21. <http://dx.doi.org/10.18596/jotcsa.83708>
- [19] Matin, M.M., & Ibrahim, M. (2010). Synthesis of some methyl 4-O-octanoyl- α -L-rhamnopyranoside derivatives. *Journal of Applied Sciences Research*, 6(10), 1527–1532. ISSN: 1816-157X
- [20] Gumel, A.M., Annuar, M.S.M., Heidelberg, T., & Chisti, Y. (2011). Lipase mediated synthesis of sugar fatty acid esters. *Process Biochemistry*, 46, 2079-2090. <https://doi.org/10.1016/j.procbio.2011.07.021>
- [21] Hannessian, S. (1983). Total Synthesis of Natural Products: The 'Chiron' Approach. Pergamon Press (Oxford). ISBN: 978-0080307152
- [22] Dhavale, D.D., & Matin, M.M. (2005). Piperidine homoazasugars: Natural occurrence, synthetic aspects and biological activity study. *ARKIVOC*, 2005(3), 110-132. ISSN: 1424-6376
- [23] Kinnaert, C., Daugaard, M., Nami, F., & Clausen, M.H., (2017). Chemical synthesis of oligosaccharides related to the cell walls of plants and algae. *Chemical Reviews*, 117, 11337–11405. <https://doi.org/10.1021/acs.chemrev.7b00162>
- [24] Kabir, A.K.M.S., & Matin, M.M., (1994). Regioselective acylation of a derivative of L-rhamnose using the dibutyltin oxide method. *Journal of the Bangladesh Chemical Society*, 7(1), 73-79. ISSN: 1022-016X

- [25] Komatsu, K., Tsuda, M., Tanaka, Y., Mikami, Y., & Kobayashi, J. (2005). SAR studies of brasilicardin A for immunosuppressive and cytotoxic activities. *Bioorganic & Medicinal Chemistry*, 13(5), 1507-1513. <https://doi.org/10.1016/j.bmc.2004.12.029>
- [26] Kim, S.R., & Kim, Y.C. (2000). Neuroprotective phenylpropanoid esters of rhamnose isolated from roots of *Scrophularia buergeriana*. *Phytochemistry*, 54(5), 503-509. [https://doi.org/10.1016/s0031-9422\(00\)00110-2](https://doi.org/10.1016/s0031-9422(00)00110-2)
- [27] Grond, S., Langer, H.J., Henne, P., Sattler, I., Thiericke, R., Grabley, S., et al. (2000). Secondary metabolites by chemical screening, 39 acyl α -L-rhamnopyranosides, a novel family of secondary metabolites from *Streptomyces* sp.: Isolation and biosynthesis. *European Journal of Organic Chemistry*, 2000, 929-937. [https://doi.org/10.1002/\(SICI\)1099-0690\(200003\)2000:6%3C929::AID-EJOC929%3E3.0.CO;2-U](https://doi.org/10.1002/(SICI)1099-0690(200003)2000:6%3C929::AID-EJOC929%3E3.0.CO;2-U)
- [28] Hu, J.F., Wunderlich, D., Sattler, I., Hartl, A., Papastavrou, I., Grond, S., et al. (2000). New 1-Oacyl α -L-rhamnopyranosides and rhamnosylated lactones from *Streptomyces* sp., inhibitors of 3 α -hydroxysteroid-dehydrogenase (3 α -HSD). *Journal of Antibiotics*, 53, 944-953. <https://doi.org/10.7164/antibiotics.53.944>
- [29] Kabir, A.K.M.S., Matin, M.M., Bhuiyan, M.M.R., Rahim, M.A., & Rahman, M.S. (2005). Biological evaluation of some monosaccharide derivatives. *International Journal of Agriculture & Biology*, 7(2), 218-221. ISSN 1560-8530
- [30] Platel, R., Chaveriat, L., Le Guenic, S., Pipeleers, R., Magnin-Robert, M., Randoux, B., Trapet, P., Lequart, V., Joly, N., Halama, P., Martin, P., et al. (2021). Importance of the C12 carbon chain in the biological activity of rhamnolipids conferring protection in wheat against *Zymoseptoria tritici*. *Molecules*, 26, 40. <https://dx.doi.org/10.3390/molecules26010040>
- [31] Matin, M.M., Ibrahim, M., & Rahman, M.S. (2008). Antimicrobial evaluation of methyl 4-O-acetyl- α -L-rhamnopyranoside derivatives. *Chittagong Univ. J. Biol. Sci.*, 3(1&2), 33-43. <http://dx.doi.org/10.3329/cujbs.v3i1.13404>
- [32] Kabir, A.K.M.S., & Matin, M.M. (1997). Regioselective monoacylation of a derivative of L-rhamnose. *J. Bangladesh Acad. Sci.*, 21(1), 83-88.
- [33] Matin, M.M., & Ibrahim, M. (2006). Synthesis of 2,3-di-O-substituted derivatives of methyl 4-O-acetyl- α -L-rhamnopyranoside. *Chittagong Univ. J. Sci.*, 30(2), 67-76. ISSN 1561-1167
- [34] Krishnamoorthy, M., & Balakrishnan, R., (2014). Docking studies for screening anticancer compounds of *Azadirachta indica* using *Saccharomyces cerevisiae* as model system. *J. Nat. Sci. Biol. Med.*, Vol.5, No.1, 108-111. <https://doi.org/10.4103/0976-9668.127298>.
- [35] Hussain, S., Liufang, H., Shah, S.M., Ali, F., Khan, S.A., Shah, F.A., Li, J.B., & Li, S. (2022). Cytotoxic effects of extracts and isolated compounds from *Ifloga spicata* (forssk.) sch. bip against HepG-2 cancer cell line: Supported by ADMET analysis and molecular docking. *Front. Pharmacol.*, 13, 986456. <https://doi.org/10.3389/fphar.2022.986456>
- [36] Snoch, W., Wnuk, D., Witko, T., Staroń, J., Bojarski, A.J., Jarek, E., Plou, F.J., Guzik, M. (2021). In search of effective anticancer agents—Novel sugar esters based on polyhydroxyalkanoate monomers. *Int. J. Mol. Sci.*, 22, 7238. <https://doi.org/10.3390/ijms22137238>
- [37] Kumer, A., Chakma, U., Matin, M.M., Akash, S., Chando, A., & Howlader, D. (2021). The computational screening of inhibitor for black fungus and white fungus by D-glucofuranose derivatives using in silico and SAR study. *Organic Communications*, 14(4), 305-322. <https://doi.org/10.25135/agc.oc.116.2108.2188>
- [38] Matin, M.M., Bhuiyan, M.M.H., Kibria, S.M., & Hasan, M.S. (2022). Synthesis, PASS predication of antimicrobial activity and pharmacokinetic properties of hexanoyl galactopyranosides and experimental evaluation of their action against four human pathogenic bacteria and four fungal strains. *Pharmaceutical Chemistry Journal*, 56(5), 627-637. <https://doi.org/10.1007/s11094-022-02687-y>
- [39] Uzzaman, M., Hasan, M.K., Mahmud, S., Fatema, K., & Matin, M.M. (2021). Structure-based design of new diclofenac: Physicochemical, spectral, molecular docking, dynamics simulation and ADMET studies. *Informatics in Medicine Unlocked*, 25, 100677. <https://doi.org/10.1016/j.imu.2021.100677>

- [40] Kabir, A.K.M.S., Matin, M.M., Sanaullah, A.F.M., Sattar, M.A. & Rahman, M.S. (2001). Antimicrobial activities of some lyxoside derivatives. *Bangladesh J. Microbiol.*, 18(1), 89-95. ISSN: 1011-9981
- [41] James, A.A., Rahman, M.R., Huda, D., Aqlan, M.F.M., Matin, M.M., Bakri, M.K.B., Kuok, K.K., & Rahman, M.M. (2022). Synthesis and characterization of novel nano-carbon mixture from Dabai (*Canarium odontophyllum*) nutshell. *BioResources*, 17(3), 4452-4469. <https://doi.org/10.15376/biores.17.3.4452-4469>
- [42] Tazeddinova, D., Rahman, M.R., Hamdan, S.B., Matin, Bin Bakri, M.K., & Rahman, M.M. (2022). Plant based polyphenol associations with protein: A prospective review. *BioResources*, 17(4). <https://doi.org/10.15376/biores.17.4.Tazeddinova2>
- [43] Tazeddinova, D., Toshev, A.D., Abylgazinova, A., Rahman, M., Matin, M.M., Bin Bakri, M.K., & Ayan, O. (2022). A review of polyphenol and whey protein-based conjugates. *BioResources*, 17(4). <https://doi.org/10.15376/biores.17.4.Tazeddinova1>
- [44] Mehdiya, R.B.A., Shaabanb, K.A., Rebaia, I.K., Smaouia, S., Bejara, S., & Mellouli, L. (2009). Five naturally bioactive molecules including two rhamnopyranoside derivatives isolated from the *Streptomyces* sp. strain TN58. *Natural Product Research*, 23(12), 1095–1107. <https://doi.org/10.1080/14786410802362352>
- [45] Elmaidomy, A.H., Mohammed, R., Owis, A.I., Hetta, M.H., AboulMagd, A.M., Siddique, A.B., Abdelmohsen, U.R., Rateb, M.E., Sayed, K.A.E., Hassan, H.M. (2020). Triple-negative breast cancer suppressive activities, antioxidants and pharmacophore model of new acylated rhamnopyranosides from *Premna odorata*. *RSC Advances*, 10, 10584. <https://doi.org/10.1039/d0ra01697g>
- [46] Bartuzi, D., Kaczor, A.A., Targowska-Duda, K.M., & Matosiuk, D. (2017). Recent Advances and Applications of Molecular Docking to G Protein-Coupled Receptors. *Molecules*, 22(2), 340. <https://doi.org/10.3390/molecules22020340>.
- [47] Kumer, A., Chakma, U., Chandro, A., Howlader, D., Akash, S., Kobir, M.E., Hossain, T., & Matin, M.M. (2022). Modified D-glucofuranose computationally screening for inhibitor of breast cancer and triple breast cancer: Chemical descriptor, molecular docking, molecular dynamics and QSAR. *J. Chil. Chem. Soc.*, 67(3), 5623-5635. <http://dx.doi.org/10.4067/S0717-97072022000305623>
- [48] Kaewmeesri, P., Pocasap, P., Kukongviriyapan, V., Prawan, A., Kongpetch, S., & Senggunprai, L. (2022) Anti-metastatic Potential of Natural Triterpenoid Cucurbitacin B Against Cholangiocarcinoma Cells by Targeting Src Protein. *Integrative Cancer Therapies*, 21, 15347354221124861. <https://doi.org/10.1177/15347354221124861>.
- [49] Matin, P., Matin, M.M., Rahman, M.R., & Kumer, A. (2023). Synthesis, antifungal, and molecular docking studies of some new di-O-isopentanoyl glucopyranosides. *Physical Chemistry Research*, 11(1), 149-157. <https://doi.org/10.22036/PCR.2022.334577.2057>
- [50] El Aissouq, A., Chedadi, O., Bouachrine, M., & Ouammou, A. (2021). Identification of novel SARS-CoV-2 inhibitors: A structure-based virtual screening approach. *Journal of Chemistry*, 2021, 1901484. <https://doi.org/10.1155/2021/1901484>
- [51] Siddikey, F., Roni, M.A.H., Kumer, A., Chakma, U., & Matin, M.M. (2022). Computational investigation of Betalain derivatives as natural inhibitor against food borne bacteria. *Current Chemistry Letters*, 11(3), 309-320. <https://doi.org/10.5267/j.ccl.2022.3.003>
- [52] Islam, F., Rahman, M.R., & Matin, M.M. (2021). The effects of protecting and acyl groups on the conformation of benzyl α -L-rhamnopyranosides: An in silico study. *Turkish Computational and Theoretical Chemistry*, 5(1), 39-50. <https://doi.org/10.33435/tcandtc.914768>
- [53] Matin, M.M., Islam, N., Siddika, A., & Bhattacharjee, S.C. (2021). Regioselective synthesis of some rhamnopyranoside esters for PASS predication, and ADMET studies. *Journal of the Turkish Chemical Society Section A: Chemistry*, 8(1), 363-374. <https://doi.org/10.18596/jotcsa.829658>
- [54] Guan, L., Yang, H., Cai, Y., Sun, L., Di, P., Li, W., Liu, G., & Tang, Y. (2018). ADMET-score - a comprehensive scoring function for evaluation of chemical drug-likeness. *Medchemcomm*, 10(1), 148-157. <https://doi.org/10.1039/c8md00472b>.
- [55] Ali, M., Karim, M.H., & Matin, M.M. (2021). Efficient synthetic technique, PASS predication, and

- ADMET studies of acylated n-octyl glucopyranosides. *Journal of Applied Science & Process Engineering*, 8(1), 648-659. <https://doi.org/10.33736/jaspe.2823.2021>
- [56] Matin, M.M., Chakraborty, P., Alam, M.S., Islam, M.M., & Hanee, U. (2020). Novel mannopyranoside esters as sterol 14 α -demethylase inhibitors: Synthesis, PASS predication, molecular docking, and pharmacokinetic studies. *Carbohydrate Research*, 496, 108130. <https://doi.org/10.1016/j.carres.2020.108130>
- [57] Pires, D.E.V., Blundell, T.L., & Ascher, D.B. (2015). pkCSM: predicting small-molecule pharmacokinetic properties using graph-based signatures. *Journal of Medicinal Chemistry*, 58(9), 4066–4072. <https://doi.org/10.1021/acs.jmedchem.5b00104>
- [58] Daina, A., Michielin, O. & Zoete, V. (2017). SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Scientific Report*, 7, 42717. <https://doi.org/10.1038/srep42717>
- [59] Ferrer, M., Perez, G., Plou, F.J., Castell, J.V., & Ballesteros, A. (2005). Antitumour activity of fatty acid maltotriose esters obtained by enzymatic synthesis. *Biotechnol. Appl. Biochem.*, 42(1), 35–39. <https://doi.org/10.1042/BA20040122>
- [60] Zhang, W., Chakravarty, B., Zheng, F., Gu, Z., Wu, H., Mao, J., Wakil, S.J., & Quioco, F.A. (2011). Crystal structure of the human fatty acid synthase thioesterase domain with an activate site-specific polyunsaturated fatty acyl adduct. *Proc. Natl. Acad. Sci. USA*, 108(38), 15757-62. <https://doi.org/10.1073/pnas.1112334108>.
- [61] Petit-Topin, I., Fay, M., Resche-Rigon, M., Ulmann, A., Gainer, E., Rafestin-Oblin, M.-E., & Fagart, J. (2014). Molecular determinants of the recognition of ulipristal acetate by oxo-steroid receptors, *Journal of Steroid Biochemistry and Molecular Biology*, 144(B), 427-435. <https://doi.org/10.1016/j.jsbmb.2014.08.008>
- [62] Mallinger, A., Schiemann, K., Rink, C., Stieber, F., Calderini, M., Crumpler, S., et al. (2016). Discovery of Potent, Selective, and Orally Bioavailable Small-Molecule Modulators of the Mediator Complex-Associated Kinases CDK8 and CDK19. *J. Med. Chem.*, 59(3), 1078–1101. <https://doi.org/10.1021/acs.jmedchem.5b01685>
- [63] Nagai, Y., Kawano, S., Motoda, K., Tomida, M., Tatebe, C., Sato, K., & Akiyama, H. (2017). Solubility Testing of Sucrose Esters of Fatty Acids in International Food Additive Specifications. *Biological and Pharmaceutical Bulletin*, 40(3), 284-289. <https://doi.org/10.1248/bpb.b16-00738>
- [64] Kumar, H., Aggarwal, N., Marwaha, M.G., Deep, A., Chopra, H., et al. (2022). Thiazolidin-2,4-dione scaffold: An insight into recent advances as antimicrobial, antioxidant, hypoglycemic agents, mechanism of action and patents granted. *Molecules*, 27, 6763. <https://doi.org/10.3390/molecules27196763>