

SHORT COMMUNICATION

Preliminary Yeast-Based Evaluation of Medicinal Plants from Indigenous Communities in the West Coast and Northern Sabah, Malaysia, for Protein Phosphatase Inhibition

FAUZE MAHMUD^{1,2,4}, NURUL AIN ISMAIL¹, MISSA HARTYNIE UBOT¹, RICHARLES EDWARD¹, SIEW ENG HOW¹, ISLAM ADEL ABDELHAKIM⁵, NGIT SHIN LAI⁴, PING-CHIN LEE^{1,2} & JUALANG AZLAN GANSAU^{*1,2,3}

¹Biotechnology Programme, Faculty of Science and Technology, Universiti Malaysia Sabah, Kota Kinabalu, Sabah, Malaysia; ²BioAgriTech Research (BioATR) Group, Faculty of Science and Technology, Universiti Malaysia Sabah, Kota Kinabalu, Sabah, Malaysia; ³Biotechnology Research Institute, Universiti Malaysia Sabah, Kota Kinabalu, Sabah, Malaysia; ⁴Institute for Research in Molecular Medicine, Universiti Sains Malaysia, Penang, Malaysia; ⁵Pharmaceutical Department, Assiut University, Aswan, Egypt

*Corresponding author: azlanajg@ums.edu.my

Received: 24 April 2024

Accepted: 23 January 2026

Published: 30 June 2026

ABSTRACT

Sabah, Malaysia, is renowned for its rich biodiversity and multicultural heritage, creating a unique blend of ethnomedicinal knowledge. A total of 22 medicinal plants traditionally used by the Bajau, Dusun, and Rungus communities in Kota Belud, Kota Marudu, and Matunggong, Sabah, were collected through a direct ethnopharmacological approach. Crude extracts were prepared using methanol extraction and evaluated for their inhibitory activity against yeast Glc7, a homolog of human protein phosphatase 1 (PP1), which is considered a promising pharmaceutical target for diseases such as cancer, malaria, and fungal infections. A yeast-based assay was used to evaluate the PP1 inhibitory activity of the crude extracts with two strains: PAY704-1 (wild-type Glc7) and PAY700-4 (Glc7-10, a heat-sensitive mutant). The strains were incubated at 28 °C and 37 °C, with or without 1 M sorbitol as an osmotic stabilizer to assess cell wall integrity under Glc7-compromised conditions. This setup enabled a clear analysis of sample-induced inhibition. Notably, Sarah (*Glochidion* sp.) and Merabau (*Senna* sp.) exhibited selective Glc7/PP1 inhibitory activity as a zone of inhibition was observed only in strain PAY704-1 at 37 °C in the absence of 1 M sorbitol. This inhibitory activity has not been previously reported. Given the high conservation of PP1 across different organisms, these plants may serve as potential sources of new PP1 inhibitors, contributing to future drug development.

Keywords: Glc7/PP1, indigenous ethnomedicine, Sabah, yeast-based

Copyright: This is an open access article distributed under the terms of the CC-BY-NC-SA (Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License) which permits unrestricted use, distribution, and reproduction in any medium, for non-commercial purposes, provided the original work of the author(s) is properly cited.

Together with kinases, phosphatases play a crucial role in numerous essential pathways and cellular processes, including glycogen metabolism, apoptosis, and the cell cycle, by regulating reversible protein phosphorylation mechanisms. Protein phosphatase 1 (PP1) is one of the most significant phosphatases in humans. In addition to its role in regulating various cellular processes, PP1 is also crucial in cancer development, proliferation, and the adaptive response to anticancer drugs. Therefore, PP1 inhibitors represent promising targets for targeted cancer therapy (Felgueiras *et al.*, 2020). Furthermore, PP1 has been implicated as a

potential target for antiparasitic (Zeeshan *et al.*, 2021) and antifungal drugs development (Loh *et al.*, 2019). Despite its therapeutic potential, PP1 has received limited attention in drug development (Felgueiras *et al.*, 2020).

Yeast is a valuable model organism for studying human cellular processes and diseases due to its high genetic and functional similarity to humans. In yeast, PP1 is encoded as Glc7, which is homologous to human PPP1CA and PPP1CC (Hamza *et al.*, 2015). Glc7 is an essential protein involved in glycogen metabolism, meiosis, protein synthesis and

sporulation (Stark, 1996; Cannon, 2010). Using yeast strains expressing both wild-type and mutant Glc7, this study aimed to evaluate the Glc7/PP1 inhibitory activity of traditional medicinal plants used by the indigenous communities of Sabah's west coast and northern regions.

Plant samples were collected from Kampung Seri Aman, Kota Marudu, Sabah, based on the ethnomedicinal knowledge obtained through interviews with the Bajau (Kota Marudu), Rungus (Matunggong) and Dusun (Kota Belud) communities (Table 1). Different parts of healthy plants, including leaves, stems, bark, and

roots, were collected, cleaned, cut into smaller pieces, and air-dried at room temperature until a constant weight was achieved. The dried samples were then ground into a fine powder using a blender. Extraction was performed using methanol at a 1:10 ratio (1 g of sample in 10 ml of methanol). The mixtures were sonicated for 20 minutes and left at room temperature for 48 hours to ensure complete extraction. The resulting suspension was filtered through Whatman No. 1 filter paper to separate solid debris, yielding crude methanol extracts. The extracts were then dried using a concentrator, and re-dissolved in methanol at 100 mg/ml (Harborne, 1998).

Table 1. The list of plant samples collected from Kampung Seri Aman, Kota Marudu, Sabah

Sample code	Vernacular name	Species	Local use	Tested part
KM1L	Rumput Seriadi	<i>Stachytarpheta</i> sp.	Treat gastric	Leaves
KM2L	Kembung	<i>Dillenia</i> sp.	Treat haemorrhoids	Leaves
KM3L	Lunai	<i>Chromolaena</i> sp.	Stop bleeding	Leaves
KM4L	Bentiang	Unknown	Treat hypertension	Leaves
KM5L	Lingkong	<i>Lygodium</i> sp.	Treat hair loss and hypertension	Leaves
KM6L	Empalas	<i>Tertracera</i> sp.	Treat hypertension	Leaves
KM7L	Pelai'	<i>Alstonia</i> sp.	Treat mumps	Leaves
KM7S				Stem
KM8S	Wonod	Unknown	To stabilize body temperature	Stem
KM9Bs	Dungun	Unknown	Treat cancer, gastric, and skin problems	Branches
KM9L				Leaves
KM10R	Bangkau	<i>Rhizophora</i> sp.	Treat hypertension	Roots
KM10L				Leaves
KM11R	Rumput Pendul	<i>Kyllinga</i> sp.	Bath after giving birth ¹	Roots
KM12R	Tangkal	<i>Ficus</i> sp.	Treat mumps	Roots
KM13L	Uli-uli Semangat	<i>Micromelum</i> sp.	Bath after giving birth ¹	Leaves
KM14L	Pako-pako	<i>Acrostichum</i> sp.	Treat hypertension	Leaves
KM15L	Sarah	<i>Glochidion</i> sp.	Bath after giving birth ¹	Leaves
KM16L	Tangan-tangan	<i>Jatropha</i> sp.	Treat skin rashes	Leaves
KM16R				Roots
KM17L	Merabau	<i>Senna</i> sp.	Treat skin rashes and infections	Leaves
KM18L	Karamunsing	<i>Melastoma</i> sp.	Treat stomachache and poisoning	Leaves
KM18R				Roots
KM19L	Balik Angin	<i>Mallotus</i> sp.	Treat cancer ²	Leaves
KM19S				Stem
KM20B	Nangka	<i>Artocarpus</i> sp.	Treat cancer ²	Bark
KM21B	Kelempopo Batu	Unknown	Treat cancer ²	Bark
KM22S	Unknown	Unknown	Treat cancer ²	Stem

Note:

KM = Kota Marudu

L = Leaves, S = Stem, Bs = Branches, R = Roots, B = Barks

¹ = mixed for a bath after giving birth, ² = consumed together for cancer treatment

Glc7/PP1 is an essential protein phosphatase in yeast, involved in numerous processes, including the maintenance of cell wall integrity at high temperatures (Andrews & Stark, 2000). The extracts were tested for Glc7/PP1 inhibitory activity using a yeast-based assay previously

developed and validated (Andrews & Stark, 2000; Ong *et al.*, 2007). This assay enables the identification of Glc7/PP1 inhibitors by analyzing the inhibition patterns of two yeast strains: PAY704-1 (expressing wild-type Glc7/PP1), and the PAY700-4 (expressing Glc7-

10, a heat-sensitive Glc7/PP1 mutant that cannot grow at 37 °C). Inhibition of strain PAY704-1 served as the primary indicator of Glc7/PP1 inhibition, while strain PAY700-4 functioned as a reference strain to confirm the presence of a selective Glc7/PP1 inhibitor.

The inhibition of Glc7/PP1 was indicated by growth defects (zone of inhibition) in the PAY704-1 strain at 37 °C only. The inhibition of Glc7/PP1 at high temperatures compromises yeast cell wall integrity; consequently, growth defects occur. In contrast, the inhibition of Glc7/PP1 should not cause yeast growth defects at 25 °C, whether in the presence or absence of 1 M sorbitol, as cell wall stability is not compromised at low temperatures without Glc7/PP1 function (Andrews & Stark, 2000; Ong *et al.*, 2007).

To confirm the presence of a selective Glc7/PP1 inhibitor, a mutant yeast strain (PAY700-4) expressing Glc7-10, a heat-sensitive Glc7/PP1 variant, was also screened. Glc7-10 is heat-sensitive and rapidly denatures at high temperatures; therefore, strain PAY700-4 is unable to grow at 37 °C. This adverse effect can be mitigated by adding 1 M sorbitol to the medium, which stabilizes cellular osmotic pressure and compensates for compromised Glc7-10 function. Similar to strain PAY704-1, the inhibition of Glc7-10 at 25 °C, whether in the presence or absence of 1 M sorbitol, does not cause yeast growth defects. Therefore, a zone of inhibition that persists in the presence of sorbitol or affects strain PAY700-4 rules out selective Glc7/PP1 inhibition (Andrews & Stark, 2000; Ong *et al.*, 2007).

To perform the assay, a loopful of each yeast strain was cultivated in Yeast Potato Dextrose (YPD) broth (10 g/L yeast extract, 20 g/L dextrose) at 28 °C, shaken at 220 rpm for 72 hours. Then, 400 µl of each strain was inoculated into YPD agar (maintained at approximately 40 °C to prevent premature solidification) with and without 1 M sorbitol, gently swirled, and

poured into Petri dishes before solidification. Paper discs impregnated with 20 µL of plant crude extracts were placed on the solidified agar. The plates were incubated at 28 °C (permissive temperature) and 37 °C (restrictive temperature) for three days, with the zone of inhibition recorded every 24 hours (Ong *et al.*, 2007).

Based on the phenotypic observations, the crude extracts from the leaves of three plants, Lunai (*Chromolaena* sp.), Sarah (*Glochidion* sp.) and Merabau (*Senna* sp.), were found to inhibit Glc7/PP1 activity (Figure 1 and Table 2). To the best of our knowledge, PP1 inhibitory activity has not been previously reported for Sarah and Merabau. Furthermore, Sarah and Merabau were shown to selectively inhibit Glc7/PP1 activity as zones of inhibition (ZOI) were only observed in yeast strain PAY704-1 at 37 °C, in the absence of 1 M sorbitol (Table 2). However, PP1 inhibitory activity of Lunai crude extract has been previously documented (Matawali *et al.*, 2019). In addition, the leaves of Karamuning (*Melastoma* sp.) and Balik Angin (*Mallotus* sp.) exhibited toxic activity (Table 2).

Senna sp. is traditionally used for the treatment of fungal infection in the studied areas (Yusoff *et al.*, 2003). Its efficacy as an antifungal agent is also widely reported (Jeruto *et al.*, 2016; Kamilah *et al.*, 2022). In addition, *Senna* sp. and *Glochidion* sp. were previously reported for anticancer (Castro *et al.*, 2023; Sharma *et al.*, 2011) and antimalarial activities (Atanu *et al.*, 2002; Hennebelle *et al.*, 2009; Khairuddin *et al.*, 2021). Interestingly, PP1 has been identified as a potential target for drug development of these diseases (Loh *et al.*, 2019; Felgueiras *et al.*, 2020; Zeeshan *et al.*, 2021). Although the connection between PP1 inhibition and these bioactivities has yet to be established, the identification of Sarah (*Glochidion* sp.) and Merabau (*Senna* sp.) as new sources of PP1 inhibitors highlights their potential for future therapeutic applications in various diseases.

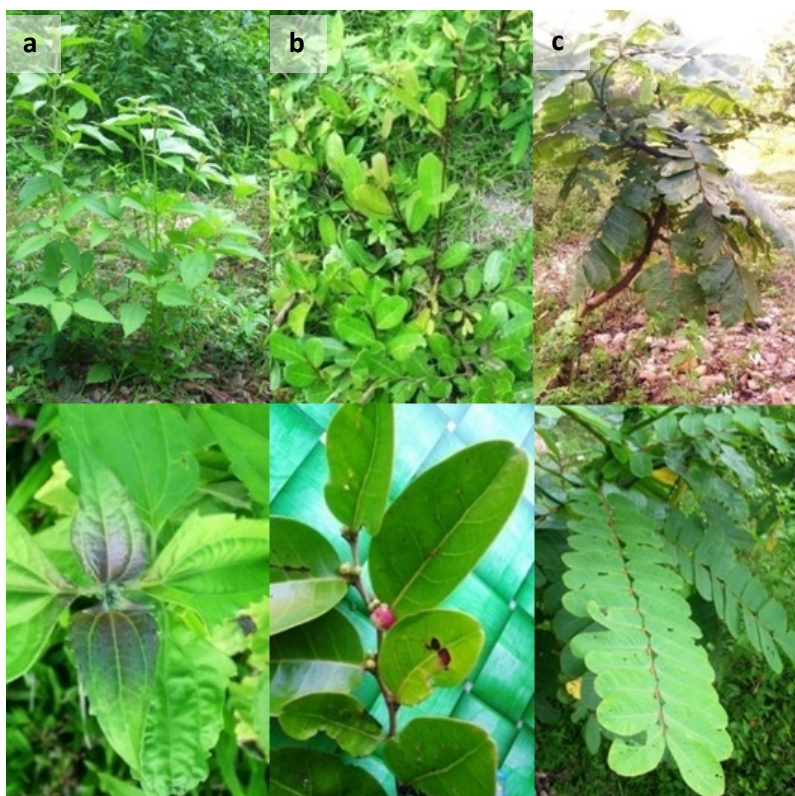


Figure 1. Traditional medicinal plants with crude leaf extracts that inhibited PP1 activity: Lunai (*Chromolaena* sp.) (a), Sarah (*Glochidion* sp.) (b), and Merabau (*Senna* sp.) (c). The top and bottom images show the whole plant and the leaves, respectively

Table 2. Inhibitory activity of plant crude extracts against Glc7/PP1 expressed in yeast cells, with KM3L (leaves of Lunai), KM15L (leaves of Sarah), and KM17L (leaves of Merabau) demonstrating inhibitory activity

Sample code	PAY700-4 (expressing GLC7-10, a heat-sensitive GLC7 mutant) (ZOI, mm)				PAY704-1 (expressing WT GLC7) (ZOI, mm)				Remarks
	YPD		YPD+1M Sorbitol		YPD		YPD+1M Sorbitol		
	25 °C	37 °C	25 °C	37 °C	25 °C	37 °C	25 °C	37 °C	
KM1L	-	-	-	-	-	-	-	-	No activity
KM2L	-	-	-	-	-	-	-	-	No activity
KM3L (Lunai)	-	-	-	-	14.00	-	-	-	Glc7/PP1 inhibitor
KM4L	-	-	-	-	-	-	-	-	No activity
KM5L	-	-	-	-	-	-	-	-	No activity
KM6L	-	-	-	-	-	-	-	-	No activity
KM7L	-	-	-	-	-	-	-	-	No activity
KM7S	-	-	-	-	-	-	-	-	No activity
KM8S	-	-	-	-	-	-	-	-	No activity
KM9Bs	-	-	-	-	-	-	-	-	No activity
KM9L	-	-	-	-	-	-	-	-	No activity
KM10R	-	No yeast growth	-	-	-	-	-	-	No activity
KM10L	-		-	-	-	-	-	-	No activity
KM11R	-		-	-	-	-	-	-	No activity
KM12R	-		-	-	-	-	-	-	No activity
KM13L	-		-	-	-	-	-	-	No activity
KM14L	-		-	-	-	-	-	-	No activity
KM15L (Sarah)	-		-	-	-	8.00	-	-	-
KM16L	-	-	-	-	-	-	-	-	No activity
KM16R	-	-	-	-	-	-	-	-	No activity
KM17L (Merabau)	-	-	-	-	8.00	-	-	-	Glc7/PP1 inhibitor
KM18L	10.00±2.83	-	-	13.00±1.41	11.00±4.25	11.00±4.25	10.00	10.00±2.83	Toxic activity
KM18R	-	-	-	-	-	-	-	-	No activity
KM19L	10.00±2.83	-	10.00±1.41	17.00±4.24	10.00±2.83	10.00	10.00±2.83	10.00±2.83	Toxic activity
KM19S	-	-	-	-	-	-	-	-	No activity
KM20B	-	-	-	-	-	-	-	-	No activity
KM21B	-	-	-	-	-	-	-	-	No activity
KM22S	-	-	-	-	-	-	-	-	No activity

Note:

KM = Kota Marudu

L = Leaves, S = Stem, Bs = Branches, R = Roots, B = Bark

Values are mean ± SD (n = 3)

ACKNOWLEDGEMENTS

This study was partially funded by research grant awarded by the Ministry of Higher Education (FRGS/1/2023/STG01/UMS/02/5). The authors would like to thank Professor Michael J. Stark (University of Dundee, Scotland), for providing the yeast strains.

REFERENCES

- Andrews, P.D. & Stark, M.J.R. (2000). Type 1 protein phosphatase is required for maintenance of cell wall integrity, morphogenesis, and cell cycle progression in *Saccharomyces cerevisiae*. *Journal of Cell Science*, 113 (Pt 3): 507–520. DOI: 10.1242/jcs.113.3.507.
- Atanu, F.O., Rotimi, D., Ilesanmi, O.B., Al Maliki, J.S., Batiha, G.E. & Idakwoji, P.A. (2002). Hydroethanolic extracts of *Senna alata* leaves possess antimalarial effects and reverses haematological and biochemical perturbation in *Plasmodium berghei*-infected mice. *Journal of Evidence-Based Integrative Medicine*, 27 (2515690X221116407). DOI: 10.1177/2515690X221116407.
- Cannon, J.F. (2010). Function of protein phosphatase-1, Glc7, in *Saccharomyces cerevisiae*. *Advances in Applied Microbiology*, 73: 27–59. DOI: 10.1016/S0065-2164(10)73002-1.
- Castro, D.T.H., Leite, D.F., da Silva Baldivia, D., dos Santos, H.F., Balogun, S.O., da Silva, D. B., Carollo, C.A., de Picoli Souzam K. & dos Santos, E.L. (2023). Structural characterization and anticancer activity of a new anthraquinone from *Senna velutina* (Fabaceae). *Pharmaceuticals*, 16 (7): 951. DOI: 10.3390/ph16070951.
- Felgueiras, J., Jerónimo, C. & Fardilha, M. (2020). Protein phosphatase 1 in tumorigenesis: is it worth a closer look? *Biochimica et Biophysica Acta - Reviews on Cancer*, 1874 (2): 188433. DOI: 10.1016/j.bbcan.2020.188433.
- Hamza, A., Tammperre, E., Kofoed, M., Keong, C., Chiang, J., Giaever, G., Nislow, C. & Hieter, P. (2015). Complementation of yeast genes with human genes as an experimental platform for functional testing of human genetic variants. *Genetics*, 201 (3): 1263–1274. DOI: 10.1534/genetics.115.181099.
- Harborne, J.B. (1998). *Phytochemical Methods - A Guide to Modern Techniques of Plant Analysis* 5th Edition. London: Chapman and Hall.
- Hennebelle, T., Weniger, B., Joseph, H., Sahpaz, S. & Bailleul, F. (2009). *Senna alata*. *Fitoterapia*, 80 (7): 385–393. DOI: 10.1016/j.fitote.2009.05.008.
- Jeruto, P., Arama, P.F., Anyango, B., Akenga, T., Nyunja, R. & Khasabuli, D. (2016). *In vitro* antifungal activity of methanolic extracts of different *Senna didymobotrya* (fresen.) H.S. Irwin & Barnaby plant parts. *African Journal of Traditional, Complementary and Alternative Medicines*, 13 (6): 168–174. DOI: 10.21010/ajtcam.v13i6.24.
- Kamilah, D., Elya, B., Adawiyah, R. & Silvyana, A.E. (2022). *Senna siamea* hexane extract: Potent antifungal activity against *Candida albicans*, *Candida krusei* and identification of its chemicals content. *Pharmacognosy Journal*, 14 (6): 999–1004. DOI: 10.5530/pj.2022.14.203.
- Khairuddin, Taebe, B. & Kombong, O.I.N. (2021). Measurement of specific and non-specific parameters of Sampare leaves ethanol extract (*Glochidion* sp var. Biak) as a traditional antimalarial agent. *Journal of Pharmaceutical and Medicinal Sciences*, 6 (1). DOI: 10.32814/jpms.v6 i1.123.
- Loh, J.T., Xu, S., Huo, J.X., Kim, S.S.Y., Wang, Y. & Lam, K.P. (2019). Dok3-protein phosphatase 1 interaction attenuates Card9 signaling and neutrophil-dependent antifungal immunity. *Journal of Clinical Investigation*, 129 (7): 2717-2729. DOI: 10.1172/JCI126341.
- Ong, S.M., Voo, L.Y.C., Lai, N.S., Stark, M.J.R. & Ho, C.C. (2007). Screening and characterization of microbial inhibitors against eukaryotic protein phosphatases (PP1 and PP2A). *Journal of Applied Microbiology*, 102 (3): 680–692. DOI: 10.1111/j.1365-2672.2006.03135.x.

- Sharma, J.V.C., Chandra, B., Chakraborty, R. & Chanda, H. (2011). Anticancer activity of aqueous extract of roots of *Glochidion zeylanicum* (Gaertn.). *Journal of Pharmaceutical and Biomedical Sciences*, 6 (11): 1–4.
- Stark, M.J.R. (1996). Yeast protein serine/threonine phosphatases: Multiple roles and diverse regulation. *Yeast*, 12 (16): 1647–1675. DOI: 10.1002/(SICI)1097-0061(199612)12:16%3C1647::AID-YEA71%3E3.0.CO;2-Q.
- Yusoff, M.M., Ahmad, B. & Pasok, G. (2003). Traditional medicinal plants of the Dusun Tobilung of Kampong Toburon, Kudat, Sabah, Malaysia. *Borneo Research Bulletin*, 34 (1954).
- Zeeshan, M., Pandey, R., Subudhi, A.K., Ferguson, D.J.P., Kaur, G., Rashpa, R., Nugmanova, R., Brady, D., Bottrill, A.R., Vaughan, S., Brochet, M., Bollen, M., Pain, A., Holder, A.A., Guttery, D.S. & Tewari, R. (2021). Protein phosphatase 1 regulates atypical mitotic and meiotic division in *Plasmodium* sexual stages. *Communications Biology*, 4 (1): 760. DOI: 10.1038/s42003-021-02273-0.